

A**MADS-box**

NsMADS1	MGRGRVQLKRIENKINRQVTF SKRASGLLKKAEI SVLCDAEVGLIVFSTKGLFEY
POTM1-1	*****R*****
SCM1	*****R*****
BpMADS5	*****R*****A*****
MdMADS2	*****R**M*****A*I*****
BpMADS3	*****AV***H*****
Saapl	*****RA*****A*V***H*****
BoAP1	*****RA**F*****A*V***H*****
AtAP1	*****RA*****A*V***H*****
CAL	*****E*****RT*****Q*****S*****H*****
SQUA	***K*****RG*****L*****A*****N*****

B**K-box**

NsMADS1	LEHAKLKARLEVLQRNQRHYAGEDLDSLSMKELQNLHQLD SALKHIRSRKNQLMHESIS ELQKK
POTM1-1	*****K*V***E**N*****V***Q
SCM1	**N*****EKL*V***E**N*****A*****V***Q
BpMADS5	*****K*FV*****L*****Q*****Y*****R*
MdMADS2	*****V*****M*****Q*****V*Y*****
BpMADS3	M*F*R**GKV*L***H***L*D**E**H*****Q**T***V*T***Y***Q***
Saapl	**YNR***KI*L*E*****L***QAM*P*****Q**T*****D**N***R*
BoAP1	M*YNR***KI*L*E*****L***QAM*P*****Q**T*****Y**N***R*
AtAP1	M*YNR***KI*L*E*****L***QAM*P*****Q**T*****T*****Y**N***R*
CAL	M*YSR***KI*L*E*****L**E*EPM*L*D*****Q**ET*****N**LNH***R*
SQUA	**YS*****I*L***H***M*****M*L*I*S*Q**T***N**T***LYD*****H*

C**C-terminal end**

NsMADS1	YPTAG-----DNGEVEG--SSRQQQ--QNTVMPPWMLRHLNG
POTM1-1	*QNTNV---V*****GN**Q**GAAN*****Q***RHLN*
SCM1	SQNTNV---V*****GN**QX*GAAN*****Q**VRHLN*
BpMADS5	-----SQAR*NGR-VDEGTPPHRA*ALL***RHLNQ
MdMADS2	SNYQAIRSEGIP*DNQYGDPTPHRP*MLL*A*IVRHLNE
BpMADS3	L*CLNI-----G*NYQ-EEAPEVRRNELELTLE*IYSC**GCFAT
Saapl	S*FLNM-----G*LYQEEDPMEMRRNDLDSLE*VYNCN*GCFAA
BoAP1	S*FLNM-----G*LYQEEDQMAMRRNDLDSLE*VYNCN*GSFAA
AtAP1	S*FLNM-----G*LYQEEDPMAMR-NDLELTLE*VYNCN*GCFAA
CAL	S*FLNM-----G*LYQ*EDQTAMRRNLDLTL* IYNY-*GCYAA
SQUA	F*CVN-----GNTY**EGANEDRRNELDLTLDLSLYSC**GCFAA

Figure 2. Multiple alignment of the amino acid sequences deduced from NsMADS1 and other MADS proteins in the SQUA subfamily. The MADS-box (A), K-box (B), and C-terminal end (C) regions were aligned. Gaps (hyphens) were introduced for the maximum sequence homology. The asterisks (*) indicate identical amino acids to NsMADS1.

present in developing fruit. The results in Figure 4 demonstrate that the *NsMADS1* transcript is present in flowers, especially in sepals and carpels, but not in vegetative organs such as leaves, stems, and roots of mature plants and in seedlings. A similar expression pattern was observed in a rice MADS gene, *OsMADS1*, if palea/lemma were regarded as the first whorl (Chung et al., 1994).

The functional role of the *NsMADS1* gene was investigated by ectopically expressing the gene in transgenic tobacco plants. The cDNA clone was put under control of the CaMV 35S promoter, and the

chimeric molecule (pGA1577) was introduced into tobacco plants by the *Agrobacterium*-mediated transformation technique (An et al., 1988). Among the 10 transgenic plants tested, we did not observe any phenotypic abnormalities (data not shown). RNA blot analysis showed that the *NsMADS1* transcript was detectable in leaves from all of the transgenic plants. Thus, this method was unable to reveal the functional role of the *NsMADS1* gene. It is possible that NsMADS1 may need another organ-specific factor to control the expression of the target gene (Chung et al., 1998). Further studies are required to understand

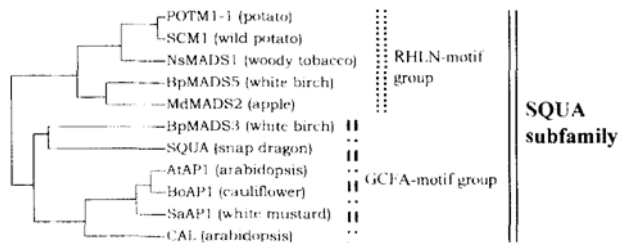


Figure 3. Phylogenetic tree for NsMADS1 and other MADS proteins in the SQUA subfamily. Shown is a neighbor-joining method tree based on the amino acid sequences.

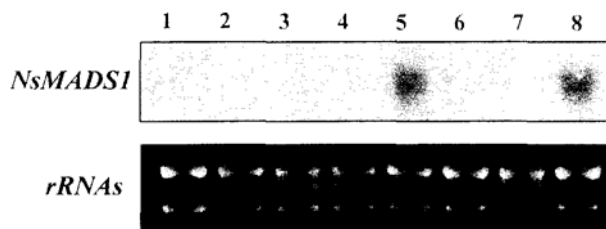


Figure 4. Expression of the *NsMADS1* gene in different organs of tobacco plants. Twenty-five micrograms of total RNA was loaded on each lane. The RNA blot was hybridized with ^{32}P -labelled *NsMADS1* probe, which was prepared from a cDNA fragment lacking the MADS-box region. 1, 15-day-old seedlings; 2, mature stems; 3, mature leaves; 4, roots from mature plants; 5, sepals; 6, petals; 7, stamens; 8, carpels. Ethidium bromide staining of rRNAs is shown to ensure equal amounts of RNA loading.

the function of the *NsMADS1* gene.

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